

Correspondence to:

Chad P. McHugh
USAFSAM/EKEN
Brooks AFB, TX
78235-5301

submitted to:

Journal of Parasitology

DEP FILE COPY

AD-A220 916

RESEARCH NOTE . . .

LEISHMANIASIS IN TEXAS: ISOLATION OF Leishmania mexicana
FROM Neotoma micropus

Chad P. McHugh, Max Grogl^{*}, and Sara F. Kerr⁺

DTIC
ELECTE
APR 24 1990
S B D
Co

Epidemiology Division, U.S. Air Force School of Aerospace Medicine, Brooks Air Force Base, Texas 78235-5301, ^{*}Division of Experimental Therapeutics, Walter Reed Army Institute of Research, Washington, D.C., 20307 and ⁺Witte Museum, 3801 Broadway, San Antonio, Texas, 78209

DISTRIBUTION STATEMENT 1

Approved for public release
Distribution Unlimited

90 04 20 004

REPORT DOCUMENTATION PAGE

Form Approved
OMB No 0704-0188

1a. REPORT SECURITY CLASSIFICATION Unclassified			1b. RESTRICTIVE MARKINGS		
2a. SECURITY CLASSIFICATION AUTHORITY			3. DISTRIBUTION/AVAILABILITY OF REPORT Approved for public release; distribution is unlimited.		
2b. DECLASSIFICATION/DOWNGRADING SCHEDULE					
4. PERFORMING ORGANIZATION REPORT NUMBER(S) USAFSAM-JA-90-12			5. MONITORING ORGANIZATION REPORT NUMBER(S)		
6a. NAME OF PERFORMING ORGANIZATION USAF School of Aerospace Medicine (USAFSAM)		6b. OFFICE SYMBOL (if applicable) EKEN		7a. NAME OF MONITORING ORGANIZATION	
6c. ADDRESS (City, State, and ZIP Code) Human Systems Division (AFSC) Brooks AFB TX 78235-5301				7b. ADDRESS (City, State, and ZIP Code)	
8a. NAME OF FUNDING/SPONSORING ORGANIZATION USAF School of Aerospace Medicine		8b. OFFICE SYMBOL (if applicable) EKEN		9. PROCUREMENT INSTRUMENT IDENTIFICATION NUMBER	
8c. ADDRESS (City, State, and ZIP Code) Human Systems Division (AFSC) Brooks AFB TX 78235-5301				10. SOURCE OF FUNDING NUMBERS	
PROGRAM ELEMENT NO.		PROJECT NO. SUPT		TASK NO. XX WORK UNIT ACCESSION NO. EK	
11. TITLE (Include Security Classification) Leishmaniasis in Texas: Isolation of <u>Leishmania mexicana</u> from <u>Neotoma micropus</u> .					
12. PERSONAL AUTHOR(S) McHugh, Chad P.; Grogg, Max; and Kerr, Sara F.					
13a. TYPE OF REPORT		13b. TIME COVERED FROM _____ TO _____		14. DATE OF REPORT (Year, Month, Day) 1990	
15. PAGE COUNT					
16. SUPPLEMENTARY NOTATION					
17. COSATI CODES			18. SUBJECT TERMS (Continue on reverse if necessary and identify by block number)		
FIELD	GROUP	SUB-GROUP			
06	03				
06	05		Leishmania mexicana, cutaneous leishmaniasis, isolation, culture, host, woodrat, Texas, Neotoma micropus.		
19. ABSTRACT (Continue on reverse if necessary and identify by block number) A female <u>Neotoma micropus</u> infected with <u>Leishmania</u> was collected in Zavala County, Texas, on January 15, 1990. The infection was limited to lesions at the bases of the ears, and the parasite grew readily in Schneider's Drosophila medium supplemented with 20% fetal bovine serum. Isozyme analysis determined the parasite to be <u>L. mexicana</u> .					
20. DISTRIBUTION/AVAILABILITY OF ABSTRACT <input checked="" type="checkbox"/> UNCLASSIFIED/UNLIMITED <input type="checkbox"/> SAME AS RPT <input type="checkbox"/> DTIC USERS			21. ABSTRACT SECURITY CLASSIFICATION Unclassified		
22a. NAME OF RESPONSIBLE INDIVIDUAL Chad P. McHugh, MPH, PhD			22b. TELEPHONE (Include Area Code) (512) 536-3471		22c. OFFICE SYMBOL USAFSAM/EKEN

✓ 2 2 2
ABSTRACT: A female Neotoma micropus infected with Leishmania was collected in
Zavala County, Texas, on January 15, 1990. The infection was limited to
lesions at the bases of the ears, and the parasite grew readily in Schneider's
Drosophila medium supplemented with 20% fetal bovine serum. Isozyme analysis
determined the parasite to be L. mexicana. *Keywords: parasite disease, (K)*



Accession For	
NTIS GRA&I	<input checked="checked" type="checkbox"/>
DTIC TAB	<input type="checkbox"/>
Unannounced	<input type="checkbox"/>
Justification	
By	
Distribution/	
Availability Codes	
Dist	Avail and/or Special
A-1	

INDEX DESCRIPTORS:

Leishmania mexicana, cutaneous leishmaniasis, Neotoma
micropus, host, woodrat, Texas, isolation, culture

The leishmaniasis, a complex of parasitic diseases, are a serious public health problem in endemic areas of Africa, Asia, the Mediterranean region, Central and South America, and southern Mexico. Reports of cutaneous leishmaniasis in humans in southern Texas and adjacent Mexico indicate that an enzootic focus also exists in this region, but the vertebrate reservoir(s) and sand fly vector(s) remain unknown (Grimaldi et al., 1989). We report the first isolation of Leishmania from a rodent host collected in Texas.

In late June, 1989, an elderly, female resident of Zavala County, Texas, was diagnosed as having cutaneous leishmaniasis. Although confirmed histologically, the parasite was not isolated in culture, and a specific identification was not made. The woman had lived at her present residence for the previous 3 years and rarely left except to attend a nearby church.

Extensive live-trapping for small mammals was conducted around the case residence in an effort to collect potential hosts of Leishmania. When trapping at the residence proved unsuccessful, the traps were relocated to a site approximately 2 km from the residence where rodent activity was apparent. Emphasis was given to collecting specimens of Neotoma micropus, the southern plains woodrat. This cricetid rodent was considered a likely zoonotic host since 1) its range (Davis, 1974) matched the distribution of human leishmaniasis cases in Texas, 2) Lutzomyia anthophora, a sand fly which has transmitted Leishmania mexicana under laboratory conditions (Endris et al., 1984), commonly was found in N. micropus nests (Young, 1972), 3) cricetid rodents are hosts for L. mexicana in other endemic areas (Lainson and Shaw, 1979), and 4) the senior author had seen evidence of woodrats within 200 m of 4 other case residences.

On January 15, 1990, 3 female N. micropus were collected. Two females (weights 180 g and 230 g) were apparently normal with no demonstrable

Leishmania infection. The pinnae of both ears on the remaining female (220 g) were completely eroded, with multiple, irregularly-bordered lesions covering the cauliflower-like swellings at the bases of the ears (Fig. 1). Numerous Leishmania amastigotes were seen in impression smears of lesion tissue stained with either Giemsa or genus-specific fluorescent-labeled monoclonal antibodies. Only 1 amastigote was seen in 200 fields of a fluorescent-stained impression smear of liver. Giemsa-stained impression smears and cultures of liver and spleen tissue were negative for Leishmania.

The primary isolation of parasites was made from a biopsy sample placed in Schneider's *Drosophila* medium (GIBCO Laboratories Inc., Grand Island, New York) supplemented with 20% (v/v) heat-inactivated fetal bovine serum (GIBCO), 500 units/ml penicillin, and 500 ug/ml streptomycin. The isolate (MNEO/US/90/WR972) was passed in the supplemented Schneider's medium until sufficient promastigotes were available for isoenzyme analysis.

Isozyme patterns were determined by cellulose acetate electrophoresis (Kreutzer and Christensen, 1980). The isolate was characterized twice for up to 20 enzymes including the 3 enzymes known to accurately identify parasites of the genus Leishmania: glucose phosphate isomerase (GPI-EC 5.3.1.9), mannose phosphate isomerase (MPI-EC 5.3.1.8), and phosphogluconate dehydrogenase (6PDGH-EC 1.1.1.44) (Kreutzer et al., 1987). The specific identification was based on comparison with World Health Organization reference strains which were run in parallel.

The apparent restriction of the parasites to the ears, the ease of isolation and rapidity of growth in culture, and its sensitivity to temperatures > 37 C are all suggestive of L. mexicana. Isozyme characterization determined that the parasite isolated from N. micropus collected in Texas is L. mexicana.

Additional studies to detail other biological and biochemical characteristics of the isolate are under way.

If N. micropus is a zoonotic reservoir of Leishmania in Texas, L. anthophora is a likely candidate for vector, at least among woodrats. Based on her weight, the age of the infected female was estimated to be 4-5 months (Raun, 1966). This estimate and the condition of the lesions suggest that transmission of Leishmania occurred in the area during the fall of 1989. Transmission in September through November is consistent with the seasonal activity patterns of L. anthophora (Addis, 1945). Other vertebrates such as opossums, hispid cotton rats, and armadillos and other sand flies such as Lutzomyia diabolica and Lutzomyia texana also may be involved in the enzootic cycle in Texas, but humans are undoubtedly accidental hosts. Additional studies to quantify the prevalence of Leishmania infections in Neotoma populations, document the involvement of L. anthophora, identify other reservoirs, determine the vector of Leishmania to humans, and detail the factors which lead to spillover of Leishmania into human populations are under way.

The authors thank Drs. D. Dimmit, B. L. Limmer and J. C. Barnes for bringing cases to our attention. Mr. T. Long and Dr. D. Spalinger arranged access to sampling sites. Dr. R. D. Kreutzer confirmed the electrophoresis results. The photograph for Figure 1 was taken by A. Holguin. This note is dedicated to L. J. McHugh who shared our enthusiasm for the search and the excitement of the discovery.

LITERATURE CITED

- ADDIS, C. J. 1945. Phlebotomus (Dampfomyia) anthophorus, n. sp., and Phlebotomus diabolicus Hall from Texas (Diptera: Psychodidae). *Journal of Parasitology* 31: 119-127.
- DAVIS, W. B. 1974. The mammals of Texas. Bulletin no. 41. Texas Parks and Wildlife Department, Austin, Texas, 294 p.
- ENDRIS, R. G., D. G. YOUNG, AND P. V. PERKINS. 1987. Experimental transmission of Leishmania mexicana by a North American sand fly, Lutzomyia anthophora (Diptera: Psychodidae). *Journal of Medical Entomology* 24: 243-247.
- GRIMALDI, G. Jr., R. B. TESH, and D. McMAHON-PRATT. 1989. A review of the geographic distribution and epidemiology of leishmaniasis in the New World. *American Journal of Tropical Medicine and Hygiene* 41: 687-725.
- KREUTZER, R. D., and H. A. CHRISTENSEN. 1980. Characterization of Leishmania spp. by isozyme electrophoresis. *American Journal of Tropical Medicine and Hygiene* 29: 199-208.
- KREUTZER, R. D., N. SOURATY, AND M. E. SEMKO. 1987. Biochemical identities and differences among Leishmania species and subspecies. *American Journal of Tropical Medicine and Hygiene* 36: 22-32.
- LAINSON, R., AND J. J. SHAW. 1979. Chapter 1. The role of animals in the epidemiology of South American leishmaniasis. In *Biology of the Kinetoplastida*, W. H. R. Lumsden and D. A. Evans (eds.). Academic Press, London and New York, pp. 1-116.
- RAUN, G. G. 1966. A population of woodrats (Neotoma micropus) in southern Texas. Bulletin no. 11. Texas Memorial Museum, Austin, Texas, 62 p.
- YOUNG, D. G. 1972. Phlebotomine sand flies from Texas and Florida (Diptera: Psychodidae). *Florida Entomologist* 55: 61-64.

FIGURE 1. Leishmania-infected Neotoma micropus collected in Zavala County, Texas, on January 15, 1990.

